

INORGANIC BORANOPHOSPHATE SALTS

FIELD OF THE INVENTION

The present invention relates to inorganic boranophosphate salts, that are
5 phosphate mimic, and to their preparation and uses.

BACKGROUND OF THE INVENTION

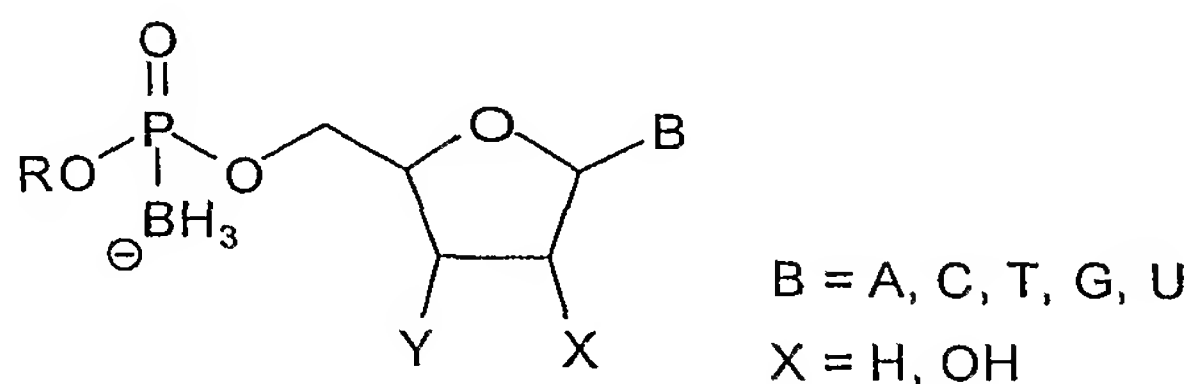
The quest for phosphate bioisosters over the last several decades included the
synthesis of phosphonates, α -halo (e.g. difluoromethyl) phosphonates (Blackburn,
1981; Blackburn *et al.*, 1981 and 1987), phosphorothioates (Nahorski and Potter,
10 1989; Eckstein, 1983, 1985, and 2000) and boranophosphate analogues (Sood *et al.*,
1990; Summers *et al.*, 1998; Shaw *et al.*, 1993 and 2000).

Phosphates and phosphate-containing molecules play a major role in
numerous biological systems (Westheimer, 1987 and 1992). However, the
unwanted lability of the ester P-O bond has promoted the search for suitable
15 bioisosters, phosphate analogues, which retain biological activity but possess
diminished lability. The search for bioisosters was initiated by the need to produce
phosphate probes for various studies, such as probing stereochemical requirements
of enzymes (Roumaniuk and Eckstein, 1981; Conolly and Eckstein, 1982). In
addition, phosphate bioisosters have been developed for improving the
20 pharmacological effects of nucleotide-based drugs, e.g. anti-sense agents (Agrawal,
1999; Stein, 1996).

A widely used isoster of phosphate is phosphorothioate and its analogues,
proposed in the pioneering work of Eckstein *et al.* (Nahorski and Potter, 1989;
Eckstein, 1983, 1985, and 2000). In these analogues, the nonbridging oxygen atom
25 is replaced by a sulfur atom. Other chemical modifications of the phosphate moiety
include the substitution of the labile phosphate ester oxygen atom by carbon or
nitrogen atom, to give phosphonates and phosphoramidate analogues, respectively
(Engel, 1977).

During the last decade, pioneering studies by Spielvogel and Ramsay-Shaw have proposed boranophosphate analogues **1** as bioisosters of natural nucleotides (Sood *et al.*, 1990; Shaw *et al.*, 2000) and as important tools for biochemists (Rait *et al.*, 1999; Zhang *et al.*, 1997; Porter *et al.*, 1997).

5



- 1a. Y = OH; R = H
 1b. Y = OH; R = $P_2O_6^{3-}$
 1c. Y = OH; R = PO_3^{2-}
 1d. Y = OH; R = 3'-nucleoside
 1e. Y = OH; R = 3'-oligonucleotide
 1f. R = 3'-ribose

This new class of boron modified nucleotides, that mimic phosphodiester, phosphorothioate, and methyl phosphonate, was designed for use as potential therapeutic and diagnostic agents. These nucleoside boranophosphates, or borane phosphonates, have a borane moiety (BH_3) in replacement of one of the nonbridging oxygen atoms in the phosphate diester moiety. The BH_3 group maintains the negative charge of a phosphate, but it does not form classical H-bonds and does not coordinate with metal ions. This modification imparts unique characteristics to boranophosphate nucleotides and nucleic acids. The boranophosphate can be considered as a "hybrid" of three well-studied types of modified phosphates, namely, normal phosphate, phosphorothioate, and non-ionic methylphosphonate. The BH_3 group in the boranophosphates is isoelectronic with oxygen (O) in the normal phosphates, and isolobal (pseudo-isoelectronic) with sulfur (S) in phosphorothioates. The BH_3 group is isosteric with the CH_3 group in the methylphosphonates. Boranophosphates would be expected to share a number of chemical and biochemical properties with phosphorothioate and methylphosphonate analogs. Thus, boranophosphate analogues have a different charge distribution and polarity than the corresponding natural nucleotides (Shaw *et al.*, 1993).

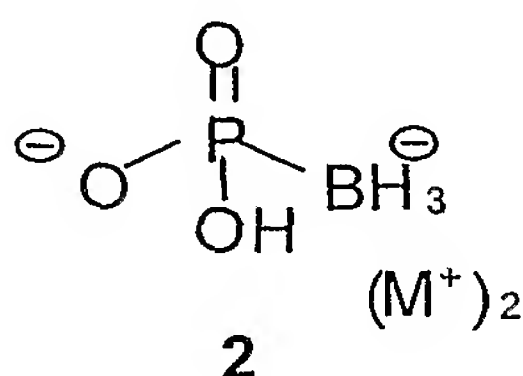
This emerging field of novel nucleotide bioisosters has expanded significantly and has provided many important applications of the boranophosphate analogues. For instance, non-terminal P-boronated nucleotides, existing as a pair of diastereoisomers, have been used as stereochemical probes to elucidate enzymatic catalysis (Sergueeva *et al.*, 2000). Oligodeoxyribonucleotides bearing boranophosphate linkages have been used for polymerase chain reaction (PCR) sequencing and DNA diagnostics (He *et al.*, 1999; Porter *et al.*, 1997), and boranophosphate nucleotides have been found to be highly potent and stable P2Y-receptor agonists (Nahum *et al.*, 2002). Oligonucleotides bearing boranophosphate linkages have also been considered as potentially useful anti-sense agents (Summers and Shaw, 2001). These analogues were also tested for the treatment of cancer as carriers of ^{10}B isotope in boron neutron capture therapy (Spielvogel *et al.*, 1992). However, despite the extensive study of related boranophosphate nucleotide/oligonucleotide analogues, the exploration of the parent inorganic boranophosphate has not been reported.

The various potential applications of a phosphate isoster, together with the limitations of the currently available isosters, justify the continued search for the perfect inorganic phosphate mimic.

20 SUMMARY OF THE INVENTION

It has now been found, in accordance with the present invention, that the inorganic boranophosphate 2, herein designated BPi, is a phosphate mimic.

The present invention thus relates to salts of the inorganic boranophosphate of the general formula 2, herein designated BPi salts, wherein M is a counterion.



In one embodiment, the counterion M is ammonium (NH_4^+) or it is an inorganic cation derived from an alkali, alkaline earth or transition metal such as, but not limited to, Na^+ , K^+ , Li^+ , Ca^{++} , Mg^{++} , Ni^{++} , Cu^{++} , Fe^{++} , Fe^{+++} , Co^{++} , Zn^{++} , Pd^{++} , and Ag^+ .

5 In another embodiment, the counterion M is an organic cation derived from an amine of the formula R_3NH^+ , wherein R is C_1 - C_{18} , preferably C_1 - C_6 , alkyl, more preferably ethyl, propyl or butyl, or two of the Rs together with the nitrogen atom to which they are attached form a 3-7 membered ring optionally containing a further heteroatom selected from the group consisting of N, S and O, such as for example
10 pyrrolidine, piperidine, morpholine, or R is phenyl or heteroaryl such as pyridyl, imidazolyl, pyrimidinyl, and the like.

The present invention further relates to a method for the preparation of BPi salt in a one-pot two-step reaction comprising reacting tris(trimethylsilyl)-phosphite with borane-dimethylsulfide complex of the formula $\text{BH}_3\cdot\text{SMe}_2$, reacting the
15 intermediate **11** (see Scheme 2 hereinafter) with the desired base in water or in methanol, thus obtaining the corresponding salt of BPi in very high yield.

In one embodiment, the intermediate **11** is treated with methanolic ammonia or with an aqueous NH_4OH solution, thus resulting in the ammonium salt of BPi, **2a**. In another embodiment, the intermediate **11** is treated with tributylamine, Bu_3N , in methanol, thus resulting in the Bu_3NH^+ salt of BPi, **2b**. In a further
20 embodiment, the intermediate **11** is treated with triethylammonium bicarbonate buffer, thus resulting in the Et_3NH^+ salt of BPi, **2c**.

In another embodiment, the compound **2a** is passed through a Sephadex-CM C-25-tetraethylammonium-form column, thus resulting in the Et_4N^+ salt of BPi, **2d**.

25 The present invention further relates to the use of the boranophosphate salts of the invention as fertilizers, in detergent formulations, as additive in melts for the glass industry, in boron neutron-capture therapy (BNCT) of cancer, and as synthetic building blocks in the synthesis of boranonucleotides that may be used for all the uses known today and that may be discovered in the future for boranonucleotides of
30 various lengths.

BRIEF DESCRIPTION OF THE FIGURES

Figs 1A-1C show the NMR spectra of BPi. **Fig. 1A:** ^1H decoupled ^{31}P NMR spectrum in D_2O at 81 MHz; **Fig. 1B:** ^1H coupled ^{31}P NMR spectrum in D_2O at 81 MHz; **Fig. 1C:** ^1H NMR spectrum in D_2O at 200 MHz.

5 **Fig. 2** shows the pH-dependent ^{31}P NMR chemical shift of BPi in H_2O within the pH range 4.87-13.20 at 81 MHz.

Figs. 3A-3B show the X-ray structure of BPi. **Fig. 3A:** Unit cell includes 8 BPi molecules, 8 H-phosphonate molecules, and 24 ammonium ions; hydrogen atoms are omitted to clarify the Bpi geometry; **Fig. 3B** ORTEP drawing of BPi;
 10 crystal data of **2a**: monoclinic, $P2_1/c$; $a=23.616(5)$ Å, $b=6.3470(13)$ Å, $c=15.325(3)$ Å; $V=2172.9(8)$ Å³; $Z=12$; $D_{\text{calcd}}=1.623$ g/cm³; $F(000)=1104$; 3094 reflections collected, $R=0.1015$, $R_w=0.2345$, $\text{GOF}=1.286$; Selected bond lengths [Å] and angles [°]: P(1)-O(1A) 1.524(7), P(1)-O(1B) 1.617(7), P(1)-O(1C) 1.583(7), P(1)-B(1) 1.891(11); O(1A)-P(1)-O(1C) 104.0(4), O(1A)-P(1)-O(1B) 105.3(4), O(1C)-
 15 P(1)-O(1B) 104.4(4), O(1A)-P(1)-B(1) 118.2(5), O(1C)-P(1)-B(1) 113.0(5), O(1B)-P(1)-B(1) 110.7(5)

Figs. 4A-4B depict the IR spectra of: (4A) BPi **2a** (KBr pellet; 1300-400 cm^{-1}), and (4B) BPi **2c** (germanium cell; 1300-1100 cm^{-1} ; cutoff: 680 cm^{-1}): curve A - methanolic solution, curve B - aqueous solution, curve C - neat sample.

20 **Fig. 5** shows determination of pKa values of BPi; plot of BPi's ^{31}P NMR chemical shift in H_2O as a function of the pH; two inflection points are observed in the pH range of 4.87-13.20.

Fig. 6 shows the ^{31}P NMR spectrum of BPi **2b** in methanolic solution.

DETAILED DESCRIPTION OF THE INVENTION

25 The present invention relates to the preparation, characterization, and unique chemical properties of inorganic boranophosphate (BPi) salts. As shown herein, the BPi ion is an excellent mimic of inorganic phosphate.

The unique and chemically interesting inorganic boranophosphate BPi was investigated here as a mimic of phosphate with respect to properties such as water
 30 solubility, geometry, acid/base character, H-bonding and chemical reactivity. The

great similarity of BPI to the inorganic phosphate, Pi, is demonstrated here by the BPI's high water solubility, and geometry that is in accordance with that of the parent, except for the long P-B bond (1.892Å) and B-P-O angles that are slightly larger than tetrahedral angles. Furthermore, the acid/base character of BPI is essentially not altered in comparison to Pi. This finding is in contrast to the corresponding phosphorothioate isoster, where there is a reduction of about two log units in the acidity relative to Pi (Jaffe and Cohn, 1978; Gerlt *et al.*, 1983). Likewise, pK_{a2} values of α-mono- and di-fluorophosphonate isosters are one and two log units, respectively, lower than pK_{a2} of phosphoric acid (Blackburn *et al.*, 1987).

BPI is stable under both highly basic and acidic conditions (at pH > 2). In addition, BPI is stable in the presence of imidazole, pyridine and divalent metal ions such as Zn²⁺ and Mg²⁺ ions. However, the P-B bond cleavage is observed upon the reaction of BPI with carbodiimides or upon catalytic hydrogenation. A loss of BOP's borane moiety also occurs at pH values below 2.

A drastic alteration in the chemical nature of BPI as compared to Pi and BH₃ complexes is observed. While Pi is a nucleophile (Saxena, 2002; El Seoud *et al.*, 2002; Cullis *et al.*, 2001; Bundgaard and Hansen, 1981), BPI is a poor nucleophile. Likewise, the reducing nature of the BH₃ group in BPI is drastically lower than in other BH₃ complexes.

The compounds 2 of the invention are all inorganic boranophosphate salts having different ammonium counterions (ammonium in 2a, tributylammonium in 2b, triethylammonium in 2c, and tetraethylammonium in 2d).

Based on the geometry, water solubility, acid-base character, and H-bonding properties, BPI appears to be perfect mimic of Pi, and is an attractive alternative to the known phosphate (thiophosphate and α-halophosphonate) isosters, and therefore, may have numerous promising applications ranging from biochemical probing to modulation of materials properties.

As mentioned above, the field of boranophosphates deals extensively with the related nucleotide/oligonucleotide analogues. However, to the best of our

knowledge, no attention has been given to the unique and chemically interesting inorganic boranophosphate 2, BPi.

The existence of BPi in the free form, $\text{BH}_3\text{O}_3\text{P}$ (CAS No. 178449-22-4), has been detected previously (Li *et al.*, 1996), while carrying out the hydrolysis of thymidine boranomonomophosphate in neutral solution. The compound was not stable: its NMR was determined in the solution, and it decomposed before it could be isolated.

Although the related dimethyl boranophosphate potassium salt 3, has been described by Imamoto *et al.* (1997) and by Wada and Saigo (Wada *et al.*, 2002), the preparation of stable salts of inorganic boranophosphate 2, has not been reported to our knowledge.



Numerous applications can be envisaged for the boranephosphates BPi, and all of them are encompassed by the present invention.

Among the essential elements required for plants growth are: P (macronutrient, 0.2 wt %) and B (micronutrient, 20 ppm). Likewise, K and N are also essential nutrients (1.0 and 1.5 wt %, respectively). Therefore, ammonium or K⁺ salt of BPi may be used in formulations of fertilizers. These salts provide the essential nutrients P, B, and N or K; they are non-acidic, water-soluble, and have a high phosphorous content.

The BPi salts of the invention can also be used for specialized detergent formulations. For this application, BPi should be provided in the form of the corresponding pyro-phosphate or tri-phosphate. Such oligoboranophosphates are expected to soften hard water by sequestering undesired Ca²⁺/Mg²⁺ ions. The high charge on the phosphate chain helps to stabilize detergent micelles (as a 'builder'). Oligoboranophosphates provide the correct pH for cleaning (slightly basic).

Furthermore, in warm/hot water ($\geq 50^{\circ}\text{C}$), boric acid is produced from hydrolysis of BPi and exerts its effect as a bleaching agent.

The BPi salts of the invention may also serve as an interesting additive in melts for glass making. For comparison, high quality borophosphate chemically durable optical glass is obtained from: $\text{MgO}/\text{Al}_2\text{O}_3/\text{K}_2\text{O}/\text{B}_2\text{O}_3/\text{P}_2\text{O}_5$ melts.

The natural boron isotope ^{10}B absorbs thermal neutrons. Upon capturing a thermal neutron, ^{10}B undergoes fission to generate ^7Li nucleus and energetic alpha (helium) particles, which are highly destructive within their relatively short path (10-14 mm). The specific localization of boron in rapidly dividing cells such as tumor cells is useful for destroying these cells by using Boron Neutron-Capture Therapy (BNCT), without affecting normal cells. BNCT requires about 5 ppm ^{10}B . Therefore, the BPi salts of the invention, which are transported to the rapidly dividing cells, can be useful as BNCT agents for treatment of tumors. The invention thus comprises the use of a boranophosphate salt as described herein for the manufacture of a pharmaceutical preparation for boron neutron capture therapy (BNCT) of cancer.

Since the BPi salts of the invention are mimics of natural phosphodiester in DNA, they can be used as synthetic building blocks for biologically active borano nucleosides and nucleotides of various lengths (mono-, di- and oligonucleotides) and designed for use as potential therapeutic and diagnostic agents, and this aspect is also encompassed by the invention. Regarding therapeutic use, we have disclosed (Nahum, 2002, WO 03/034978) that ATP- α -Boron analogues are potent $\text{P}_2\text{Y}_1\text{-R}$ (ATP receptor) agonists and can be utilized as therapeutic agents for the treatment of Type II diabetes.

The boranonucleotides obtained from the inorganic boranophosphate salts of the invention can be used in all known and future applications of borano nucleotides. For example, non-terminal P-boronated nucleotides can be used as stereochemical probes to elucidate enzymatic catalysis. The oligodeoxyborano-ribonucleotides can be used for polymerase chain reaction (PCR) sequencing and DNA diagnostics. The oligoboranonucleotides can be useful as anti-sense agents

targeting specific mRNA sequences, as inhibitors of ATP-utilizing enzymes (e.g. NTPDase) that are involved in various health disorders, and also in the treatment of cancer as carriers of ^{10}B isotope in boron neutron capture therapy.

The borano nucleotides may be prepared by any suitable synthetic method, for example as described in Sood et al. (1990), Summers and Shaw (2001) and WO 95/06752 (Shaw and Porter).

The invention will now be illustrated by the following non limiting Examples.

EXAMPLES

10 Experimental

(i) General.

All air- and moisture-sensitive reactions were performed in flame-dried, nitrogen flushed flasks sealed with rubber septa; the reagents were introduced with a syringe. The progress of the reactions was monitored by TLC on precoated Merck silica-gel plates (60 K-254). Column chromatography was performed with Merck silica gel 60 (230-400 mesh). Compounds were characterized by nuclear magnetic resonance (NMR) spectroscopy using Bruker DPX-300, DMX-600, or AC-200 spectrometers. NMR spectra were recorded with a Bruker AC-200 spectrometer with a ^{31}P NMR probe (isotope frequency of 81 MHz) using 85% H_3PO_4 as an external reference. IR spectra of BPi in KBr pellets were recorded with a Nicolet Impact 400D spectrometer using the OMNIC program. IR spectra of BPi in solution were measured using a Bruker Vector 22 equipped with a liquid nitrogen cooled MCT detector. For the ATR measurements, a Harrick variable angle ATR accessory was used. For one spectrum, 100 scans were coadded at a resolution of 4 cm^{-1} . The clean ATR Germanium crystal (Harrick Scientific Corporation) was measured for the background spectra (cutoff 680 cm^{-1}). Crystallographic data were collected with a Nonius KappaCCD diffractometer at 120K with scans of 1° collected at a speed of $1^\circ/20\text{sec}$; the merging R -factor on the data was 0.046 with 36867 reflections collected and 2979 unique. Bpi crystals were obtained as colorless needles. Further

details of the crystal structure investigation may be obtained from the Fachinformationzentrum Karlsruhe, 76344 Eggenstein-Leopoldshafen, Germany, on quoting the depository number CSD-413735. Melting points were measured using a Fisher-Johns melting point apparatus. Apparent pH values were measured with a Hanna Instruments pH-meter (HI 8521), equipped with an Orion micro-combination pH electrode (9802).

(ii) Synthesis of dibenzyl boranophosphate 9.

The synthesis was carried out according to Scheme 1B hereinafter. To a solution of dibenzylphosphite (300 μ L, 1.186 mmol) in dry THF *N*, *O*-bis(trimethylsilyl)acetamide (880 μ L, 3.56 mmol) was added with pipettor and the mixture was stirred for 25 min at room temperature. The solution was cooled to 0 °C, and 2M $\text{BH}_3\cdot\text{SMe}_2$ complex in THF (2.9 mL, 5.8 mmol) was added. The solution was stirred at room temperature for 15 min, and then evaporated. 24% NH_4OH solution (6 mL) was added and the mixture was stirred at room temperature for 1 h, and then freeze-dried. The product was purified by silica gel column chromatography (elution with CHCl_3 : MeOH, 12:1) and obtained as colorless oil in 71% yield (231 mg, 0.84 mmol). ^1H NMR (CDCl_3 , 300 MHz): δ 7.22 (s), 4.86 (m), 0.3 (1-1-1-1 quartet) ppm. ^{31}P NMR (CDCl_3 , 81 MHz): δ 97.5 (q) ppm. MS FAB (negative) m/z : 275.140 (M^-).

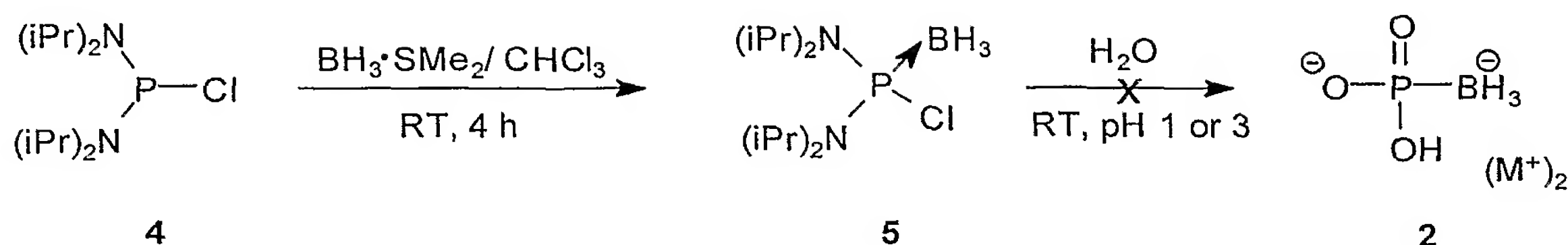
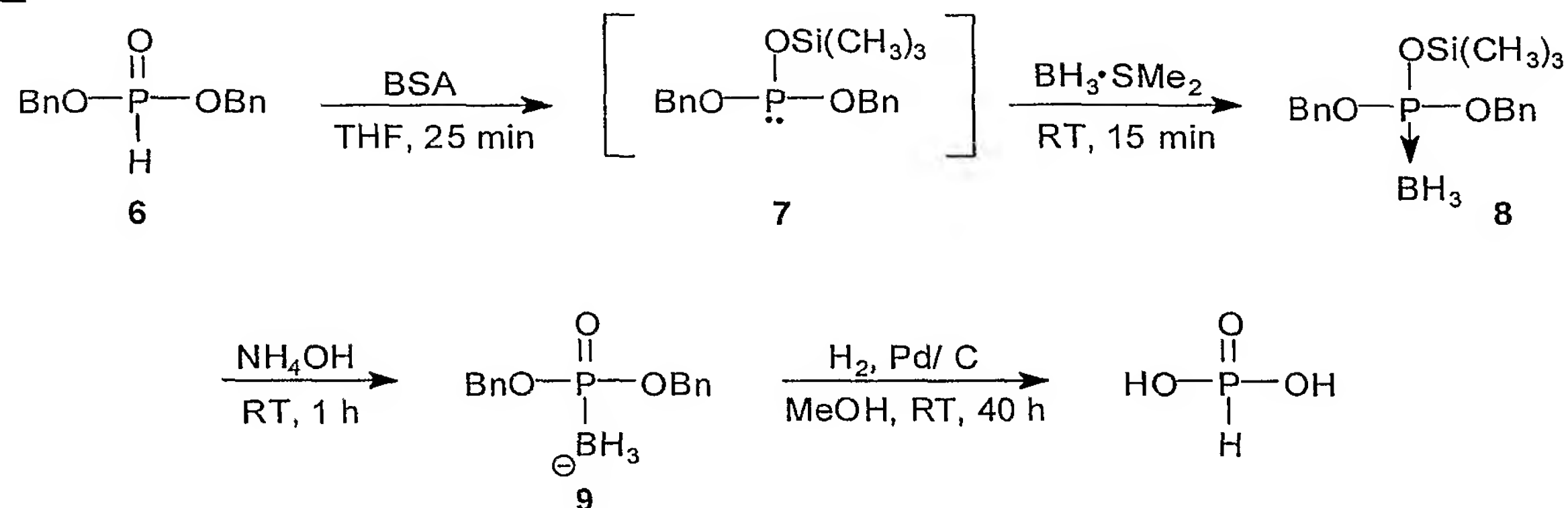
Example 1. Synthesis of BPi salts

For the preparation of boranophosphate BPi, we first attempted the treatment of chlorobis(di-isopropylamino)phosphane **4** with borane dimethylsulfide ($\text{BH}_3\cdot\text{SMe}_2$) complex (Longeau and Knochel, 1996), followed by acidic hydrolysis (pH 3 or 1) for several hours, according to Scheme 1A below. This attempt resulted in a mixture of several phosphorus species but BPi was not obtained.

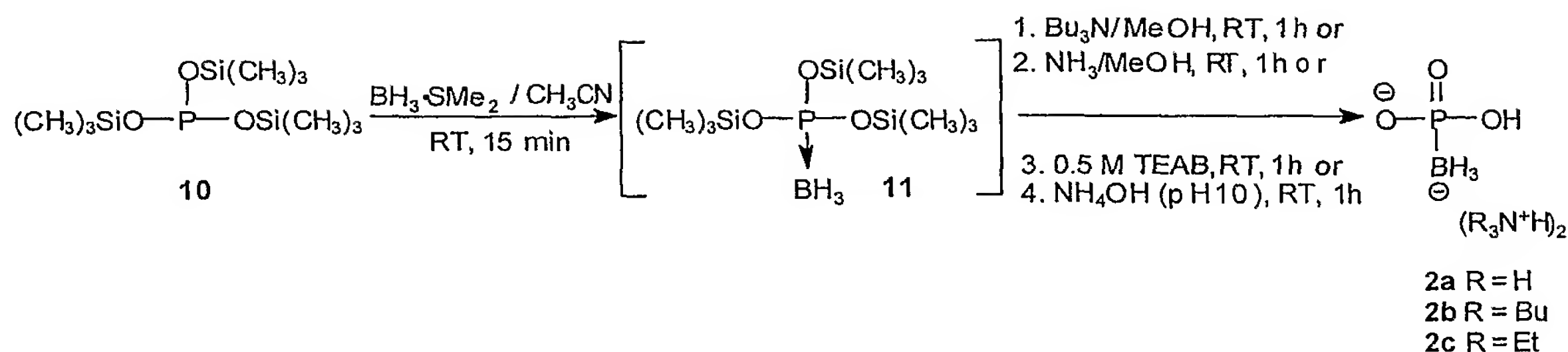
In an alternative approach, depicted in Scheme 1B below, dibenzyl H-phosphonate **6** was treated with bis(silyl)acetamide (BSA) in THF, followed by boranation of the intermediate **7** with $\text{BH}_3\cdot\text{SMe}_2$ complex, and hydrolysis of compound **8** with concentrated ammonium hydroxide for 1 h. In this way, dibenzyl

boranophosphate **9** was obtained in 71% overall yield (Scheme 1B). However, attempts to remove the benzyl groups by either catalytic hydrogenation or acidic hydrolysis (pH=1.3), resulted in the cleavage of the P-B bond, leading to phosphorus acid instead of BPi.

Scheme 1

AB

Eventually, we were able to obtain BPi in an excellent overall yield in a two-step, one-pot reaction starting from tris(trimethylsilyl) phosphite (Sood *et al.*, 1991) **10** (Scheme 2). Phosphite **10** was treated with $\text{BH}_3\cdot\text{SMe}_2$ complex in dry acetonitrile under an inert gas for 15 minutes. Subsequently, intermediate **11** was treated with 2 M methanolic ammonia for 1 h to give the ammonium salt BPi **2a**, as a white solid in 93% yield. No further purification was conducted, since volatile silyl derivatives and the unreacted $\text{BH}_3\cdot\text{SMe}_2$ were removed by evaporation. Alternatively, intermediate **11** was treated with $\text{NH}_4\text{OH}_{(\text{aq})}$ solution (pH=10), tributylamine (Bu_3N) in MeOH, or 0.5 M triethylammonium hydrogencarbonate buffer (pH=7.5) and freeze-dried or concentrated to provide the corresponding BPi salts **2a**, **2b** or **2c**, respectively.

Scheme 2

Product **2a** is highly water-soluble, whereas **2b** dissolves only in organic solvents
 5 such as MeOH, CH₃CN, DMF, and CHCl₃. Product **2c** is highly soluble both in
 water and in organic solvents.

Example 2. Synthesis and Characterization of Compounds 2a, 2b and 2d**2(i) Synthesis of ammonium boranophosphate 2a.**

10 The synthesis was carried out according to Scheme 2 hereinabove. To a
 solution of tris(trimethylsilyl)phosphite (600 μL, 1.795 mmol) in dry CH₃CN (5
 mL) under N₂ at 0 °C, BH₃·SMe₂ complex in THF (2M, 1.35 mL, 2.7 mmol) was
 added. The resulting solution was kept at room temperature for 15 min. Dry MeOH
 (15 mL) and 2 M NH₃ in EtOH (1.8 mL, 3.6 mmol) were added and the mixture
 15 was stirred at room temperature for 1 h. The solvent was removed under reduced
 pressure and the product was obtained as a white solid in 93% yield (202 mg, 1.556
 mmol), mp > 240 °C. ¹H NMR (D₂O, 200 Hz): δ 0.27 (d of 1-1-1-1 quadruplet, J_{P,H}
 = 22, J_{B,H} = 87 Hz, 3 H). ³¹P NMR (D₂O, 81 MHz): δ 80.38 (1-1-1-1 quadruplet, J =
 156 Hz, 1-1-1-1-1-1-1 septuplet, J = 52 Hz) ppm. IR (KBr): ν 2412, 2378, 2352,
 20 1181, 1149, 1077-903, 654 cm⁻¹.

2(ii) Synthesis of tetraethylammonium boranophosphate 2d.

Compound **2a** was converted to the corresponding tetraethylammonium salt
 as follows: **2a** was passed through a Sephadex-CM C-25 – tetraethylammonium-
 25 form column (prepared from the corresponding sodium form resin upon loading

with excess Et₄NCl) and the column was washed with about 20 volumes of deionized water. The solution was freeze-dried to yield tetraethylammonium BPi, **2d**, as a white solid. Based on the pH value of the **2d** solution, the ³¹P NMR spectrum, and correlation with the plot of BPi ³¹P NMR shifts vs. pH (**Fig. 5**), the expected empirical formula is BH₃O₃PH_{1.5}(Et₄N)_{1.5} (289.3): calcd. H 11.9, P 10.7; found H 11.3; P 9.5.

2(iii) Synthesis of tributylammonium boranophosphate 2b.

The tributyl ammonium salt of the inorganic boranophosphate was prepared as described above for **2a**. However, Bu₃N (0.85 mL, 3.57 mmol) was added instead of NH₃/EtOH. The product was obtained as a white solid in 93% yield (645 mg, 1.385 mmol), m.p. 83-84° C. IR (KBr): ν: 2407, 2381, 2350, 1184, 1150, 1100-850, 655 cm⁻¹.

2(iv) Determination of the pK_a values of boranophosphate 2a.

The pK_a values of **2a** were evaluated by ³¹P NMR spectroscopy at room temperature. Solutions of **2a** (0.15-0.18 M) at different pH values were prepared by adding dilute sodium hydroxide or hydrochloric acid solutions. The ³¹P NMR chemical shift was monitored as a function of the pH. A five-parameter sigmoid function was fitted to the data using Sigma Plot 2000 (SPSS, Inc.):

$$\delta = \delta_0 + a/[1 + e^{-(\text{pH}-\text{pH}_0)/b}]^c$$

The inflection point, which is determined by the second derivative of the fitted sigmoid function, is the pK_a value.

2(v) Determination of the decomposition rate of BPi 2a at pH=2.

The stability of **2a** in acidic solution was evaluated by ³¹P NMR spectroscopy at room temperature, monitoring the formation of the deboration product (phosphorus acid). A 0.16 M solution of **2a** at pH 2 was prepared by adding dilute hydrochloric acid to a solution of inorganic boranophosphate (NH₄⁺ salt) in H₂O and 10% D₂O. The percentage of decomposition of **2a** is based on integrations

of BPi and phosphorus-acid signals ($\delta = 90.93$ and 3.3 ppm, respectively). The decomposition rate was determined by measuring changes in the integration of the respective NMR signals within 96 h.

5 *2(vi) NMR spectroscopy of Compound 2a.*

Compound **2a** in water was characterized by ^{31}P NMR spectroscopy showing a signal at $\delta \approx 80$ ppm (**Fig. 1**). The boranophosphate ^{31}P NMR spectrum shows a typical pattern including two overlapping signals: the larger signal is due to coupling of P to the ^{11}B isotope, and the smaller signal is due to coupling with ^{10}B isotope. The relative height of the smaller peak to the larger one is 0.14 (**Fig. 1A**) (Harris, 1986). BPi's hydrogen- coupled ^{31}P NMR spectrum shows further splitting of the lines into a quadruplet (**Fig. 1B**). The ^1H NMR spectrum shows a typical doublet of quadruplets pattern, at $\delta \approx 0.2$ ppm, due to coupling of H to both ^{11}B and ^{31}P (**Fig. 1C**). This pattern overlaps a more complex pattern due to coupling of H to both ^{10}B and ^{31}P .

The chemical shift of BPi is pH-dependent. For instance, at pH 4.87 and 13.20 the phosphorus atom of BPi resonates at $\delta = 84$ and 63 ppm, respectively (**Fig. 2**). Likewise, the P-B coupling constant is also pH-dependent, and is reduced as the pH decreases (e.g. 147 and 183 Hz for pH 4.87 and 13.2, respectively). The pH-dependent BPi spectrum indicates structural changes of BPi, which are due to the reduction of O-P-O angles upon protonation of the molecule.

2(vii) X-ray crystallography of 2a.

To obtain structural information on BPi, compound **2a** was crystallized from an aqueous solution (pH=7). In addition to compound **2a**, the crystal contained phosphorus acid (H-phosphonate) in a 1:1 ratio. This unexpected ratio does not reflect the molar ratio in the original BPi solution, in which phosphorus acid was less than 5%.

The unit cell contains 8 BPi ions, 8 H-phosphonate ions, and 24 ammonium ions (**Fig. 3A**). Apparently, for each BPi anion, one ammonium counterion is

observed, whereas two ammonium counterions are observed around each H-phosphonate group.

For BPi, the average P-B bond length is 1.892 Å, whereas for the three P-O bonds, the average lengths are: 1.585, 1.605 and 1.524 Å, respectively (Fig. 3B). A deviation from tetrahedral angles was observed with values of 111-118 ° for B-P-O and 104-105 ° for O-P-O angles.

Comparison with X-ray crystal data obtained for the related dimethyl boranophosphate salt, **3** (Imamoto *et al.*, 1997), indicated similar values for the B-P (1.895 Å) and O-P (1.490, 1.597 and 1.612 Å) bond lengths. For dimethyl boranophosphate, one potassium ion was found near one of the oxygen atoms at a distance of 2.66 Å. Based on a comparison of the bond lengths of dimethyl boranophosphate salt with BPi, we assume that the BPi bears two H atoms, which were not found in the crystallographic data.

The shortest P-O bond (1.524 Å) indicates a partial double-bond character, and is in accordance with values found in the structures of phosphate diesters (1.47-1.51 Å) and monoesters (1.49-1.53 Å). This P-O bond is significantly longer than the bond observed in phosphate triesters (1.38-1.44 Å) (Corbridge, 1974).

2(viii) Infra-red (IR) spectroscopy of 2a and 2b.

The IR spectra of **2a** or **2b** in KBr pellet indicated characteristic bands for P-B and B-H in addition to bands associated with P-OH and P=O (Fig. 4A). Specifically, three absorptions at 2350, 2381, 2407 cm⁻¹ (*s*) (not shown) correspond to B-H stretches, and the absorption at 654 cm⁻¹ (*m*) is the P-B stretch (Corbridge, 1995). We also based our IR assignment on quantum mechanical calculations as follows. The boranophosphate anion was optimized using the B3LYP functional in conjunction with the 6-31+6(d) basis-set. Frequency calculation was performed to obtain the IR spectrum and the harmonic vibrational frequencies were scaled by a factor of 0.9614. The calculations employed the Gaussian 98 program (Frisch *et al.*, Gaussian 98 (Revision A.7), Gaussian, Inc., Pittsburgh, PA, 1998). Typical

absorptions were observed for P-OH and P=O stretches, 900-1080 cm^{-1} , and at 1140-1250 cm^{-1} , respectively.

For an evaluation of the effects of the solvents on H-bonds between BPi ions, IR spectra of BPi, **2a**, in aqueous and methanolic solutions (see below “H-bonding of Bpi”) were measured in a germanium cell and compared to the corresponding spectrum of a neat sample of Bpi (**Fig. 4B**). Comparison of those spectra indicated only minor differences. For instance, a shift of about 10 cm^{-1} to lower frequencies was observed for the P=O stretch of BPi, either in the neat sample or in MeOH, relative to BPi in aqueous solution. This shift is probably due to H-bonding based clustering in the neat sample and MeOH. The typical fine-structure for the P=O stretch in a neat sample of BPi, in the range of 1144-1178 cm^{-1} , which is possibly also due to H-bonded clusters, is lost in water. The corresponding spectrum in MeOH appears as an average of the neat sample and aqueous solution spectra, probably indicating the presence of both BPi clusters and solvent H-bonded species.

Example 3. Chemical properties of Compounds 2a, 2b and 2c

3(i) Acid-base properties.

The acid-base character of BPi was studied by ^{31}P NMR – monitored pH-titration, as described in Experimental, (v). The chemical shift of compound **2a** was plotted against pH (**Fig. 5A**). For the pH range of 4.8 - 13.2, two inflection points were observed. The second derivatives of the fitted function provided two pK_a values: 7.12 (**Fig. 5B**) and 12.54, with R^2 values of 0.999 and 0.997, respectively. These values are similar to the corresponding values of the second and third protonation equilibria of phosphoric acid (7.21 and 12.67), and are higher than those for phosphorus acid (H-phosphonate; 1.8 and 6.2).

3(ii) Stability of BPi.

BPi is stable in neutral and basic solutions. For instance, after 48 h at room temperature at pH 13.7, no degradation of BPi was observed by ^{31}P NMR spectroscopy. BPi is also relatively stable in acidic solution at pH > 2. At pH 2, BPi

slowly degrades slowly to phosphorus acid at a rate of $7 \times 10^{-7} \text{ sec}^{-1}$, $R^2 = 1.00$ ($t_{1/2} = 275 \text{ h}$), as monitored by ^{31}P NMR spectroscopy.

Under highly acidic conditions ($\text{pH} < 2$), the evolution of H_2 is clearly observed, the P-B bond is cleaved, and boric acid is formed together with
5 phosphorus acid (Scheme 3) (Li *et al.*, 1996). Phosphorus acid was observed in the ^{31}P NMR spectroscopy as a doublet at $\delta = 3.5 \text{ ppm}$ ($J = 633 \text{ Hz}$). The borane reacts with water to liberate hydrogen gas and boric acid.

The stability of inorganic boranophosphate, resulting from neutral hydrolysis of thymidine 5'-boranomonomophosphate at 50°C , has been reported earlier (Li *et al.*,
10 1996).

3(iii) *H-bonding of BPi.*

Solutions of **2b** or **2c** in organic solvents (DMF, CH_3CN , CHCl_3 and even MeOH) show unexpected ^{31}P NMR spectra. Product **2b** in MeOH apparently
15 consists of three different but pattern-related signals. The signals with chemical shifts of $\delta = 80.0$ (A), 86.2 (B) and 90.8 (C) ppm, each had an identical BPi-typical pattern (**Fig. 6**).

Several minutes after the dissolution of **2b** in MeOH, signals A, B, and C are observed in the ^{31}P NMR spectrum, with A and B as the major peaks (C constitutes
20 ca. 5% of the mixture). The composition of the initial mixture is time-dependent due to interconversion of the species. When monitoring this process with 0.14 M **2b** in CD_3OD ($\epsilon = 33$) for 160 h at room temperature, we noted the conversion of A and B to C, with a final C:B ratio of 4.4:1 (A disappeared completely). The ^1H -coupled ^{31}P NMR spectrum indicated H-splitt quadruplet signals for B and C,
25 namely, no D-H exchange occurred.

A spectrum similar to the one shown in **Fig. 6**, and time-dependent interconversion of the species, was observed for **2b** in DMF, CH_3CN , and CHCl_3 .

The possibility that the additional BPi-like species are the corresponding mono- or di-methyl esters, due to a reaction of **2b** with MeOH, was ruled out

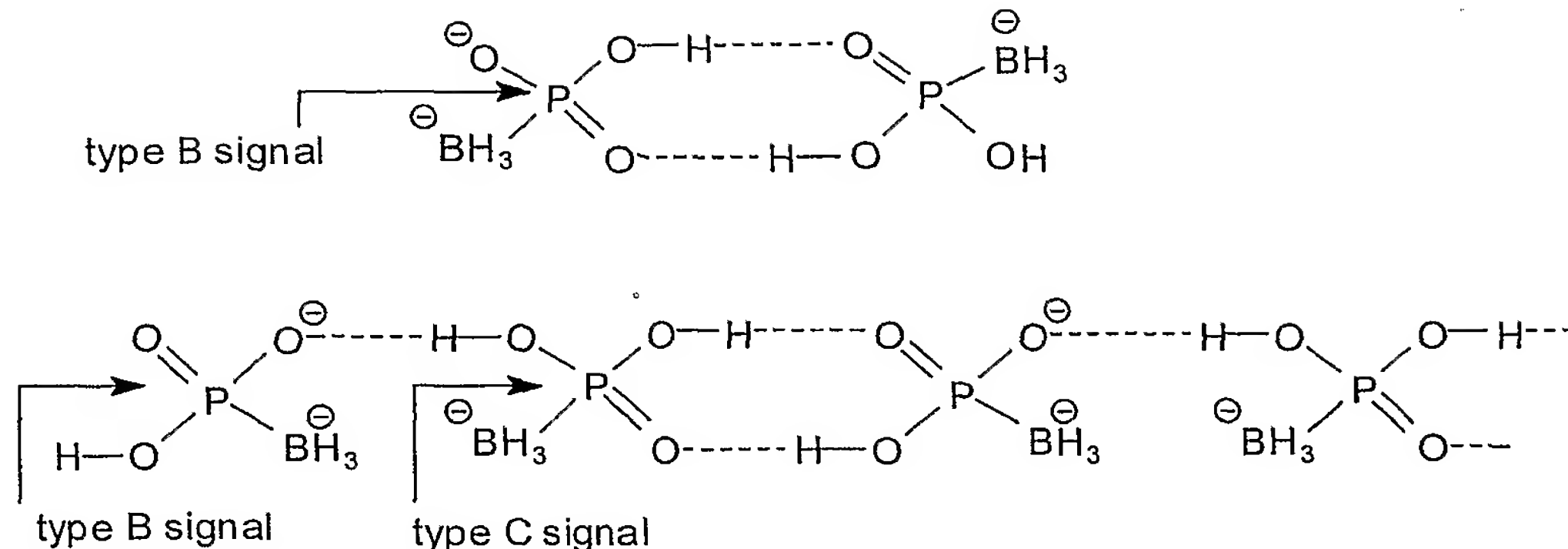
because their ^{13}C and ^1H NMR spectra in CDCl_3 were devoid of a methyl ester signal.

The three signals seen in ^{31}P NMR spectrum of **2c** in organic solvents converged into one, probably A ($\delta=79.8$ ppm), after solvent evaporation and dissolution in D_2O . Therefore, the possibility that signals B and C are due to boranophosphate anhydrides, resulting from **2c** in the NMR sample is unlikely.

To assess the possibility of observing different H-bond-clustered species on the NMR timescale, we measured the ^{31}P NMR spectrum of the parent phosphate bis(tributylammonium) salt in benzene, where clustering is known to occur (Peppard *et al.*, 1958; Peppard *et al.*, 1957). Indeed, three signals were clearly observed at $\delta=3.63$, 3.23, and 2.93 ppm, demonstrating that different H-bonded phosphate clusters can be detected by ^{31}P NMR spectroscopy. These three phosphate signals, seen in benzene, converged into one in acetonitrile and MeOH, indicating the collapse of the Pi clusters in polar/protic solvents.

Based on our observations of the pH-dependent chemical shift of BPi (**Fig. 2**), and on the determination of BPi's acidity constants (**Fig. 5**), we propose the following assignment of signals A, B and C. Signal A corresponds to the monomeric BPi, whose chemical shift at $\delta=80$ ppm indicates that half the BPi monomer population bears two protons, and the other half bears one proton (**Fig. 5**). Signal B, at $\delta=86$ ppm, corresponds to a BPi moiety that has one BPi H-bonded neighbor. Namely, signal B could result both from BPi dimers and higher clusters (Scheme 4). In these cases, each BPi is associated with an additional proton (Bu_3NH^+ ions neutralize the negative charges). Therefore, the chemical shift of the BPi dimer shifts downfield ($\delta=86$ ppm, as at pH 4.7). As indicated by signal C, BPi also forms clusters, corresponding to a BPi moiety that has two H-bonded BPi neighbors (Scheme 4). A BPi moiety in the middle of a cluster is associated with three protons, resulting in an additional downfield shift to $\delta=91$ ppm, corresponding to that of BPi at pH=2.

Scheme 4



Small H-bonded clusters (i.e. dimers and trimers) are formed almost
 5 instantaneously. This is probably the stage of nucleation. Once a critical nucleus is
 formed, a slow process of high-order clustering occurs. At this stage the
 concentration of A in solution is drastically reduced. This H-bonding based
 clustering mechanism is also supported by the observation that, upon evaporation of
 the organic solvent from the species mixture and dissolution in water, only A is
 10 detected.

The fact that BPI forms clusters even in MeOH, whereas Pi forms clusters
 only in benzene, implies that BH₃ may play a role in the pre-organization of the BPI
 clusters. The lipophilic BH₃ moieties possibly form the core of the cluster due to
 hydrophobic interactions (in MeOH). This core is then further stabilized by P-
 15 O⁻...HO-P H-bonds.

3(iv) Reactions of BPI with selected reagents.

The reactivity of BPI towards various organic and inorganic reagents was
 explored as part of the characterization of BPI's chemical nature. These reagents
 20 include: aqueous acid solution, nitrile, amide, carbodiimide, pyridine and imidazole,
 tosyl chloride, phosphorous oxychloride, H₂, and Zn²⁺ and Mg²⁺ ions.

Although BH₃, in complexes with a variety of sulfur/amine/oxygen
 compounds, is an efficient reducing agent, its reducing nature is drastically altered
 in Bpi. For instance, while hydrid transfer from "BH₃" to water occurs readily, the

BH₃ moiety in BPi transfers hydride only in a highly acidic solution (pH<2). Likewise, while BH₃·THF complex readily reduces nitriles and amides to the corresponding amines (Brown, 1972), the borane moiety in BPi does not reduce acetonitrile and dimethyl formamide, as evidenced by the complete stability of BPi
5 in these solutions.

A carbodiimide reagent is used for the condensations of phosphate to provide the corresponding phosphoric anhydride. The reaction of **2a** with an excess of 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide (EDC) was explored in water (pH 6.5), at 37°C for 4 h. The addition of EDC to BPi resulted in excessive loss of this
10 compound, due to complete P-B bond cleavage of **2a**, to yield phosphorus acid (72 % of **2a** was degraded after 4 h, based on the ³¹P NMR spectrum). This finding is in contrast to diethylphosphite (cyano- or methoxycarbonyl)borane analogues, which are stable to dicyclohexylecarbodiimide (DCC) (Vyakaranam *et al.*, 2002).

The P-B bond was also found to be sensitive to catalytic hydrogenation.
15 Thus, when compound **9** was subjected to hydrogenation (over Pd/C), the P-B bond was also reduced, yielding phosphorus acid.

The reactivity of BPi with imidazole and pyridine was studied. Specifically, a solution of BPi with 2 or 10 equiv. of imidazole in CD₃OD remained unchanged for 96 h, based on the ³¹P NMR spectra. Likewise, only a negligible cleavage of the
20 P-B bond was observed after 113 h for a solution of BPi in pyridine. BPi is apparently more stable to imidazole and pyridine than the related analogue, tetramethyl boranopyrophosphate. The reaction of 5'-DMT-2'-deoxy-thymidine with tetramethyl boranopyrophosphate in the presence of N-methylimidazole was reported to proceed with the partial removal of the borane group. Likewise, when
25 pyridine was used as a solvent, partial removal of the borane group was observed (Wada *et al.*, 2002).

The presence of divalent metal ions such as Zn²⁺ and Mg²⁺ in DMF and water for 48h and 4h, respectively, left BPi unchanged.

Whereas dimethyl boranophosphate monopotassium salt, **3**, plays the role of
30 an efficient nucleophile (Imamoto *et al.*, 1997), the related BPi is a poor

nucleophile. Thus, when BPi was treated with tosyl chloride or mesyl chloride (with or without amine) in acetonitrile for 24 h, even at 60° C, no reaction occurred. Likewise, the reaction of BPi with phosphorus oxychloride and its derivatives (P(O)Cl₂R) yielded no product.

REFERENCES

- Agrawal, S. *Biochim. Biophys. Acta* **1999**, 1489, 53-67.
- Blackburn, G. M. *Chem. Ind.* **1981**, 134.
- Blackburn, G. M.; England, D. A. Kolkman, F. *Chem. Commun.* **1981**, 930-
5 932.
- Blackburn, G. M. Brown, D.; Martin, S. J.; Parratt, M. J. *J. Chem. Soc. Perkin Trans. 1* **1987**, 181-186.
- Brown, H. C. *Borane in Organic Chemistry*. Cornell University Press: Ithaca, **1972**, pp 230-5.
- 10 Bundgaard, H.; Hansen, J. *Int. J. Pharm.* **1981**, 9, 273-283.
- Conolly, B. A.; Eckstein, F. *Biochemistry*, **1982**, 21, 6158-6167.
- Corbridge D.E.C. *The Structural Chemistry of Phosphorous*; Elsevier: Amsterdam, **1974**.
- Corbridge D.E.C. *Phosphorous: An Outline of its Chemistry, Biochemistry*
15 *and Technology*; Studies in Inorganic Chemistry, Vol 20, Elsevier: Amsterdam, **1995**; pp 1141-2.
- Cullis, P. M.; Fawcett, G. A.; Harger, M. J.; Lee, M. J. *Am. Chem. Soc.* **2001**, 123, 4147-54.
- Eckstein, F. *Ann. Rev. Biochem.* **1985**, 54, 367-402.
- 20 Eckstein, F. *Angew Chem. Int. Ed. Engl.* **1983**, 22, 423-439.
- Eckstein, F. *Antise. Nucl. Acid Drug Dev.* **2000**, 10, 117-121.
- El Seoud, O. A.; Ruasse, M.-F.; Rodrigues, W. A. *Perkin Trans. 2*, **2002**, 1053-1058.
- Engel, R. *Chem. Rev.* **1977**, 77, 349-367.
- 25 Gerlt, J. A.; Reynolds, M. A.; Demou, P. C.; Kenyon, G. L. *J. Am. Chem. Soc.* **1983**, 105, 6469-6474.
- Harris, R. K. *Nuclear Magnetic Resonance Spectroscopy: A Physicochemical View*. Longman Scientific and Technical, Somerset: NJ, 1986.
- He, K.; Porter, K. W.; Hasan, A.; Briley, J. D.; Shaw, B. R. *Nucleic Acids*
30 *Res.* **1999**, 27, 1788-1794

Imamoto, T.; Nagato, E.; Wada, Y.; Masuda, H.; Yamaguchi, A.; Uchimar, T. *J. Am. Chem. Soc.* **1997**, *119*, 9925-9926.

Jaffe, E. K.; Cohn, M. *Biochemistry*, **1978**, *17*, 652-657.

Li, H.; Hardin, C.; Ramsay Shaw, B. *J. Am. Chem. Soc.* **1996**, *118*, 6606-
5 6614.

Longeau, A.; Knochel, P. *Tetrahedron Lett.* **1996**, *37*, 6099-6102.

Nahorski, S. R.; Potter, V. B. L. *Trends Pharmacol. Sci.* **1989**, *10*, 139-144.

Nahum, V.; Ubl, J.; Reiser, G.; Levesque S.; Beaudoin, A. R.; Fischer B. *J. Med. Chem.* **2002**, *45*, 5384-5396.

10 Peppard, D. F.; Ferraro, J. R.; Mason, G. W.; *J. Inorg. Nuclear Chem.* **1958**, *7*, 231-244.

Peppard, D. F.; Ferraro, J. R.; Mason, G. W.; *J. Inorg. Nuclear Chem.* **1957**, *4*, 371-372.

Porter, K. W.; Briley, D. J.; Shaw, B. R. *Nucleic Acids Res.* **1997**, *25*, 1611-
15 1617.

Rait, V.; Sergueev, D. S.; Summers, J. S.; He, K.; Huang, F.; Krzyzanowska, B.; Shaw, B. R. *Nucleosides Nucleotides* **1999**, *18*, 1379-1380.

Roumaniuk, P.J.; Eckstein, F. *J. Biol. Chem.* **1981**, *256*, 7322-7328.

Saxena, S. Ind. J. Chem. Section A: Inorg. Bioinorg Phys. Theor. Anal.
20 Chem. **2002**, *41A*, 718-722.

Sergueeva, Z. A.; Sergueev, D. S.; Ribeiro, A. A.; Summers, J. S.; Shaw, B. R. *Helv. Chim. Acta* **2000**, *83*, 1377-1391, and references therein.

Shaw, B. R.; Sergueev, D.; He, K.; Porter, K.; Summers, J. S.; Sergueeva, Z.; Rait, V. *Methods in Enzymol.* **2000**, *313*, 226-257.

25 Shaw, B. R.; Madison, J.; Sood, A.; Spielvogel, B. F. *Methods in Molecular Biology*; **1993**; Vol. 20, Chapter 11, 225-243.

Sood, A.; Sood, C. K.; Hall, I. H.; Spielvogel, B. F. *Tetrahedron*, **1991**, *47*, 6915-6930.

Sood, A.; Ramsay Shaw, B.; Spielvogel, B. F. *J. Am. Chem. Soc.* **1990**, *112*,
30 9000-9001.

Spielvogel, B. F.; Sood, A.; Shaw, B. R.; Hall, I. H.; Fairchild, R. G.; Laster, B. H.; Gordon, C. *Prog. Neutron Capture Therap. Cancer*, **1992**, 211-213

Stein, C. A. *Chem. Biol.* **1996**, 3, 319-323.

Summers, J. S.; Roe, D.; Boyle, P. D.; Colvin, M.; Shaw, B. R. *Inorg. Chem.*
5 **1998**, 37, 4158-4159.

Summers, J. S.; Shaw, B. R. *Curr. Med. Chem.* **2001**, 8, 1147-1155.

Vyakaranam, K.; Rana, G.; Spielvogel, B. F.; Maguire, J. A.; Hosmane, N. S. *Nucleosides, Nucleotides and Nucleic Acids*, **2002**, 21, 581-598.

Wada, T.; Shimizu, M.; Oka, N.; Saigo, K. *Tet. Lett.* **2002**, 43, 4137-4140.

10 Westheimer, F. H. *Science* **1987**, 1173-1178.

Westheimer, F. H. *Phosphorus Chemistry - Developments in American Science*. Walsh, E. N.; Griffith, E. J.; Parry, R. W.; Quin, L. D. Eds. ACS Symposium Series 486ACS, Washington DC, 1992.

Zhang, J.; Terhorst, T.; Matteucci, M. D. *Tet. Lett.* **1997**, 38, 4957-4960.

15